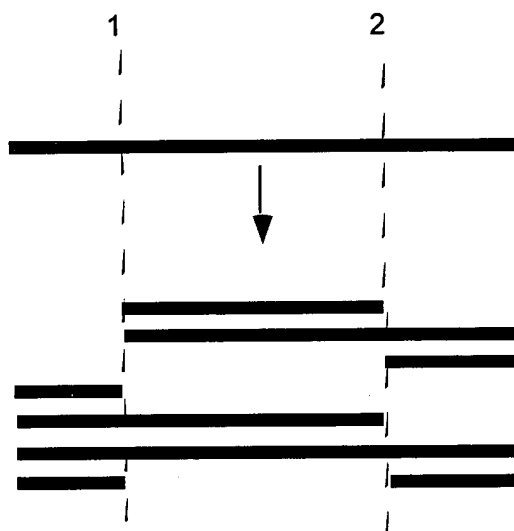


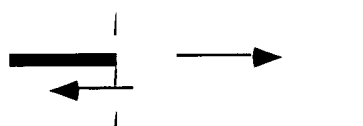
(A)

All possible recombinants  
prepared by crossover  
at positions 1 and 2



(B)

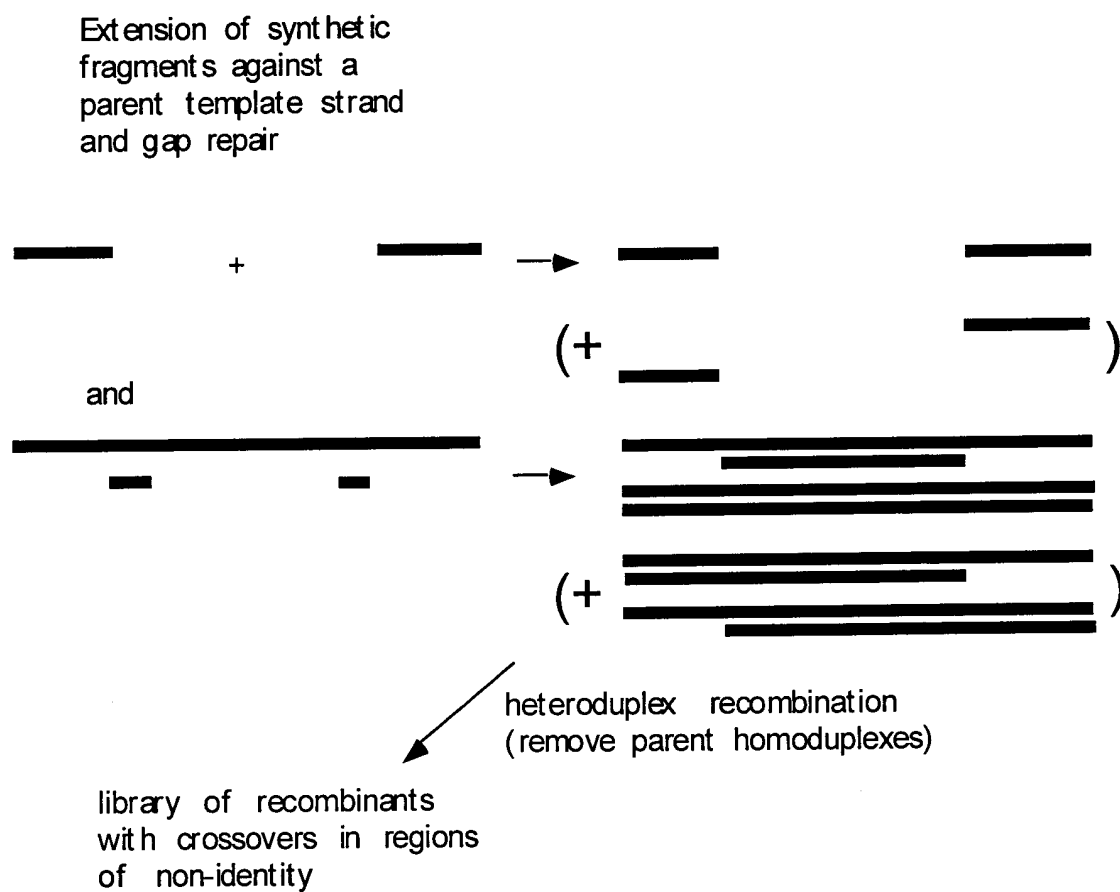
These can be prepared by  
assembly of synthetic  
fragments containing the  
crossover positions



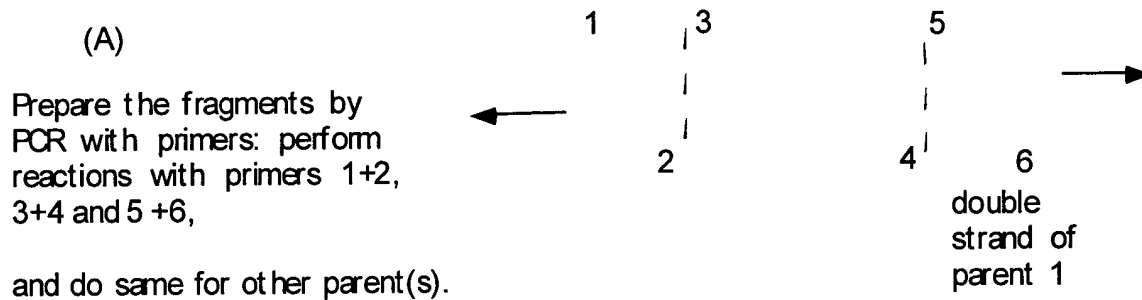
Requires fragments  
(plus end primers):



**FIG. 7**



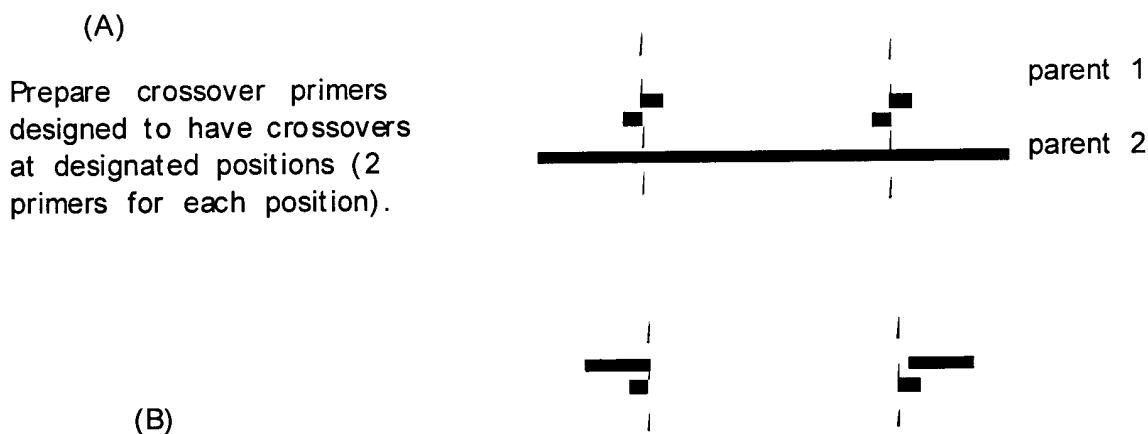
**FIG. 8**



(B)

Reassemble fragments in a pool, by PCR with 1+ 6

**FIG. 9**

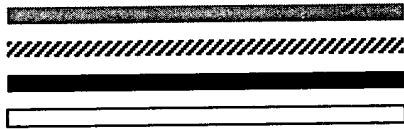


Fragment parent genes and PCR reassemble in the presence of the crossover primers to promote recombination at designated positions

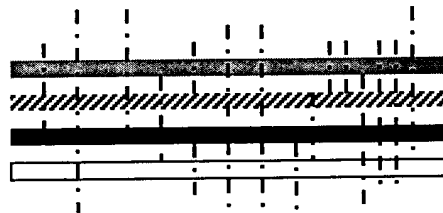
**FIG. 10**

## Recombinant search algorithm

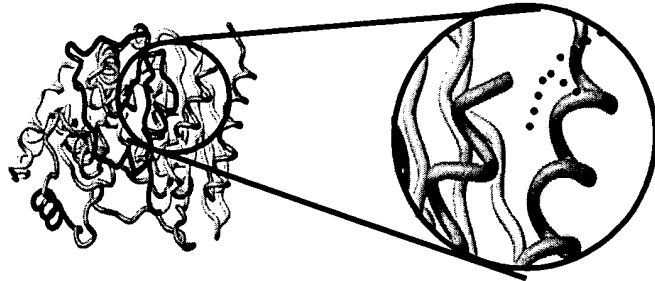
1. Align parent sequences with template structure



2. Determine all possible crossover points according to sequence identity algorithm



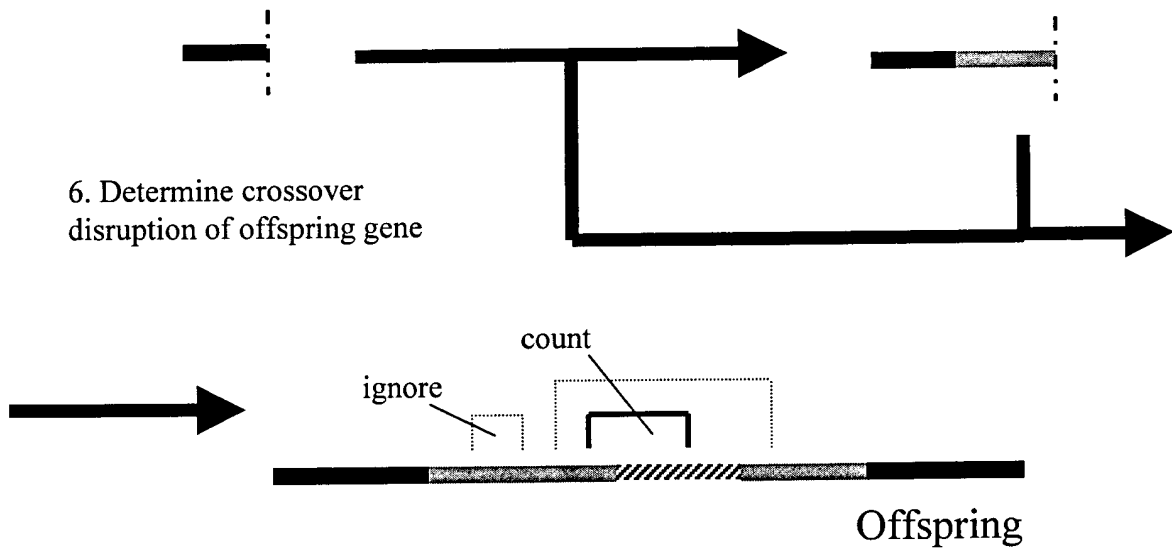
3. Calculate coupling matrix



4. Pick start parent at random and copy to offspring until a possible cut point is reached

5. Pick random number, if less than  $p$ , copy random new parent until next cut point is reached.

6. Determine crossover disruption of offspring gene



**FIG. 12**

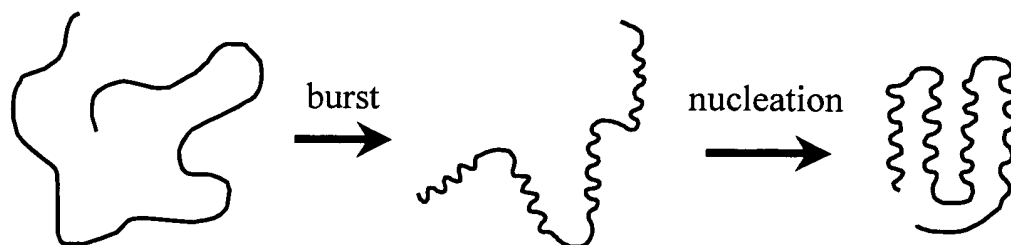


FIG. 18

The contact map shows residues that are distant (black) and residues that are close (white). If a given segment,                     , folds an above average number of residues into a given sphere size, then it is compact.

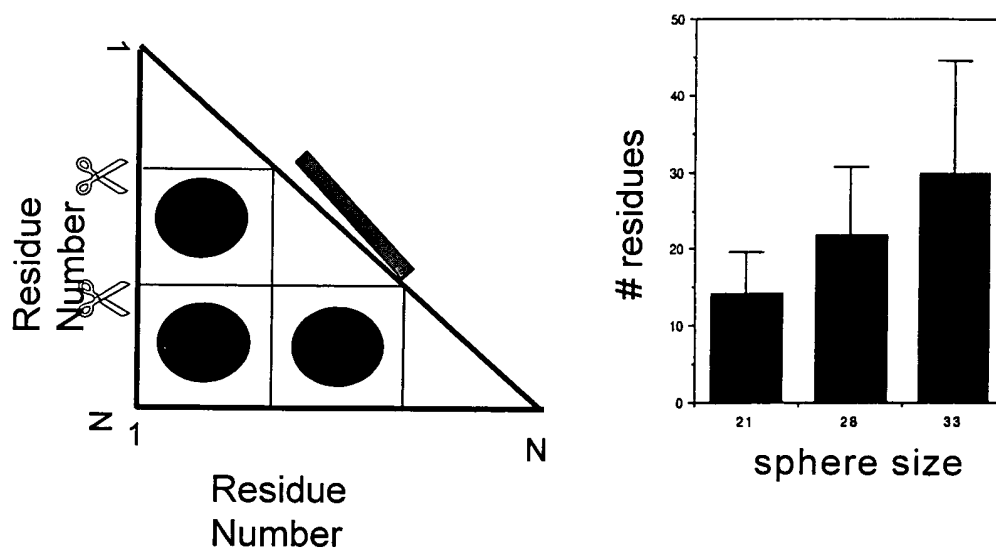
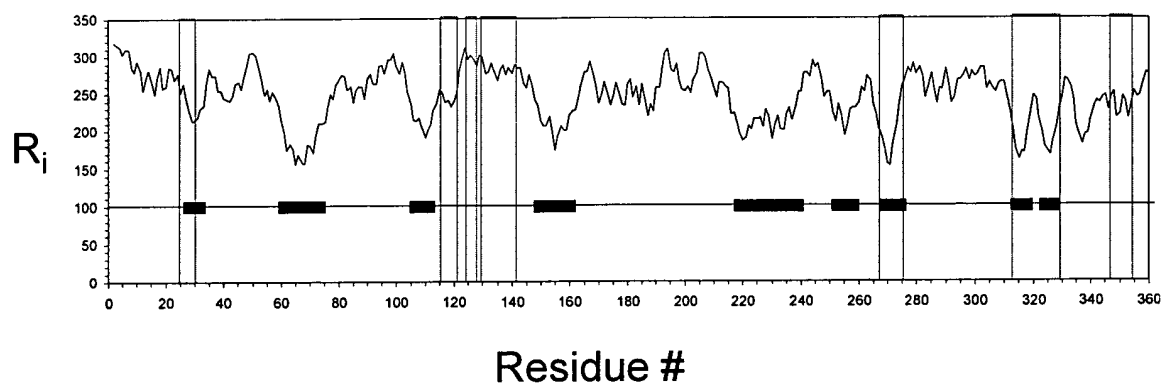
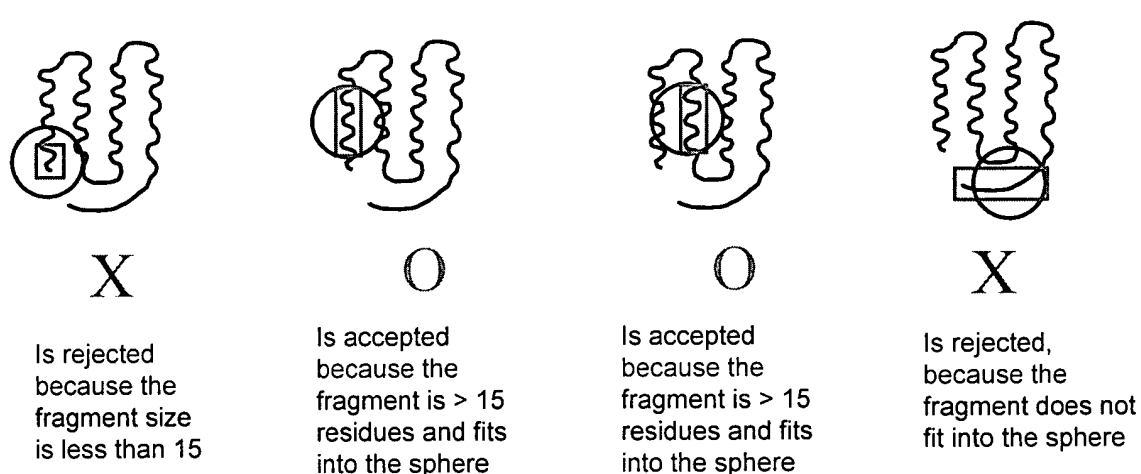


FIG. 19



**FIG. 22**



(1) Pick a sphere size (21 angstroms, like Go-Gilbert) and a disruption threshold; (2) Scan protein using segments at least the average number of residues for that sphere size or greater (e.g., >15 for 21 angstrom sphere); (3) Check the disruption of all the compact fragments identified in step 2. If the fragment has a disruption above a threshold value, keep it; otherwise, throw it out; (4) If the compact unit is disruptive, increment the schema disruption measure for all of the residues in the fragment by one. This indicates that crossovers within the fragment are disfavored.

**FIG. 23**